

Viral vectors as tools for neuroscience

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Lecture 2 (AGT):

How to make viral vectors for cell type-specific transgene expression in the brain:

- Viral vector construction
- Targeting gene expression to a specific cell type

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Physiological
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Viral Vectors for Cell Type-Specific Transgene Expression in the Brain

- Introduce the transgene into brain cells
 - Make the appropriate delivery vehicle:
 - Adenoviral vector (AVV)
 - Lentiviral vector (LVV)
- Control expression to the desired cell type



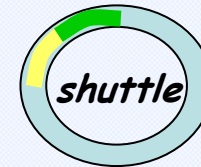
Viral Vector Production

- Shuttle plasmid (design of expression cassette)

transgene cassette



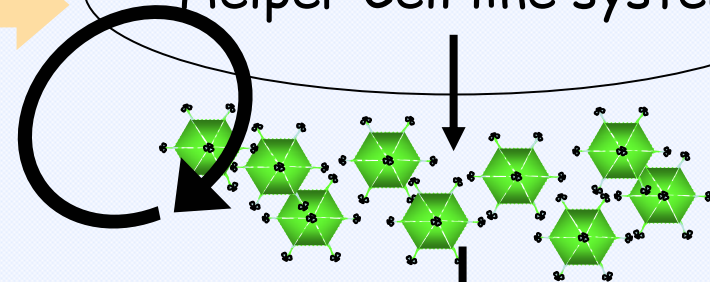
molecular cloning



- VV construction
- VV proliferation (*AVV only*)

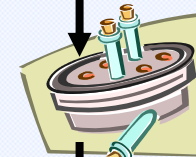
cell culture handling

Helper Cell line system



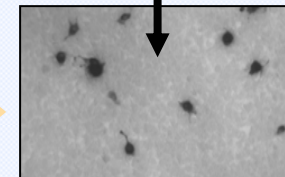
- VV purification

centrifugation and column purification



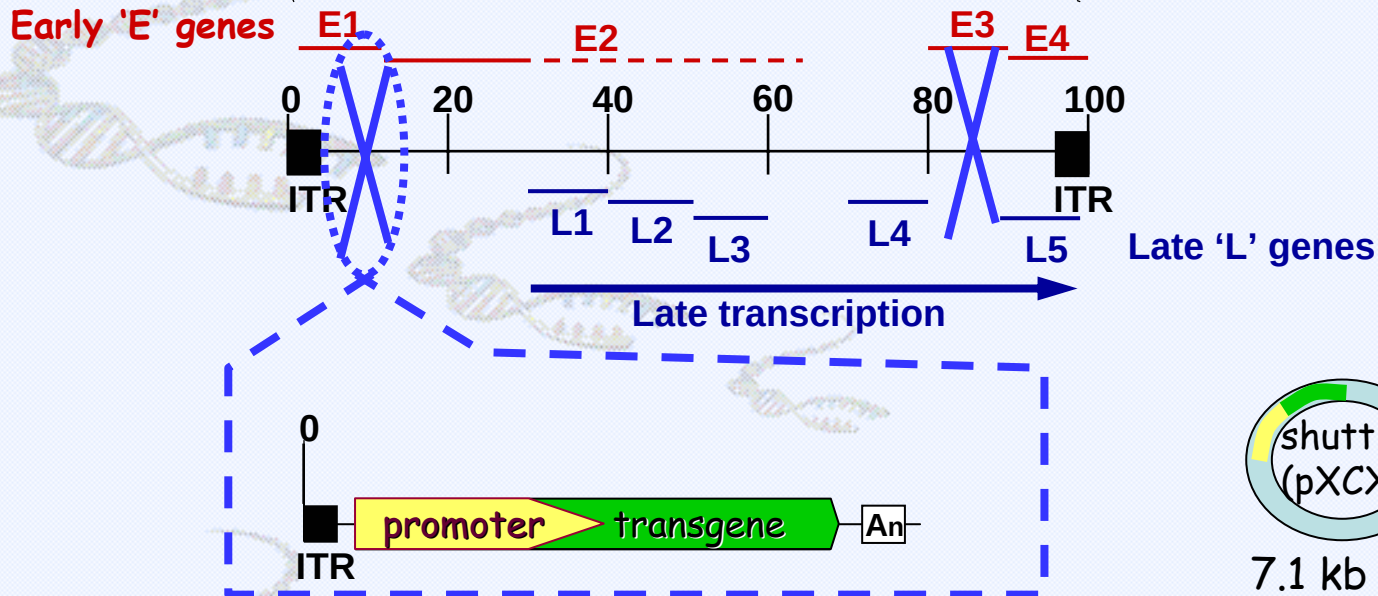
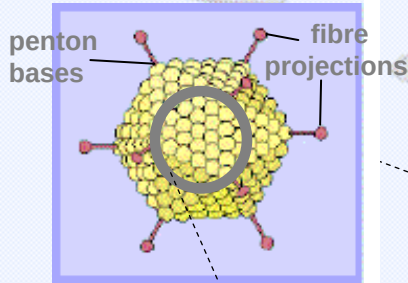
- Titer evaluation

Cell line staining & counting

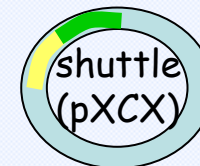


AVV - background

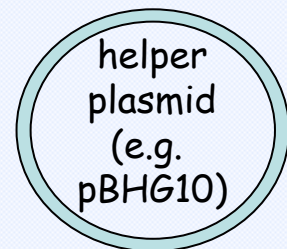
- Derived from a non-enveloped double stranded DNA virus which causes mild human infections
- Receptor-dependent uptake (CAR)
- Genome remains episomal in host cell nucleus



40 kb
wildtype
genome



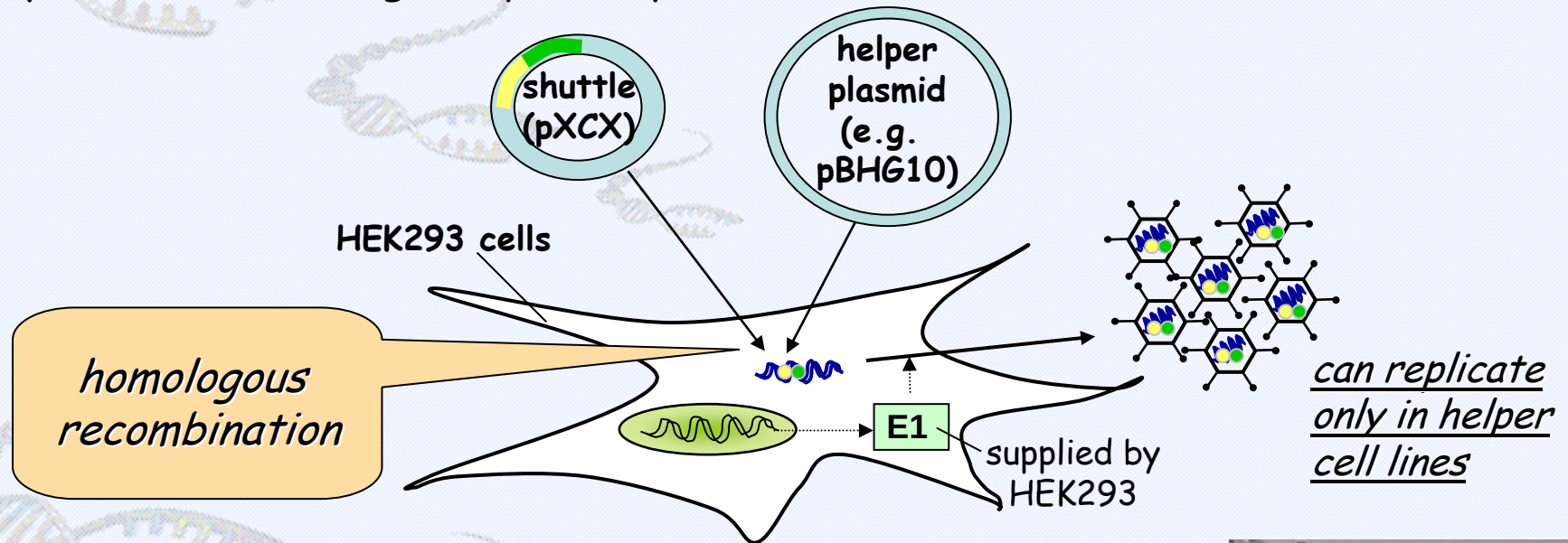
7.1 kb +
transgene
cassette



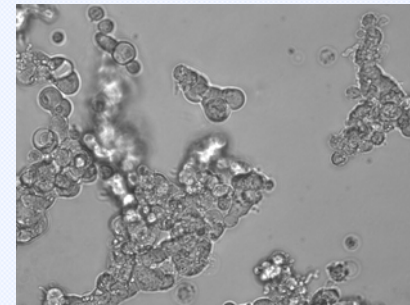
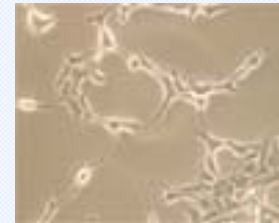
35 kb

AVV - construction & proliferation

- Co-transfection of shuttle & helper plasmid into HEK 293 cells which provide the lacking Early Gene products

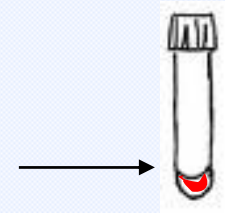
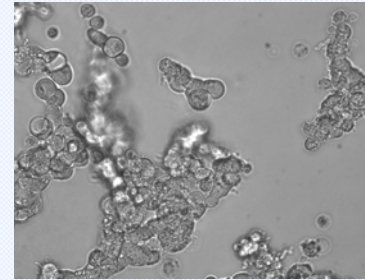


- Viral production followed by host cell death
- Enrichment steps (re-infection)
- Verification
 - Genome restriction digest analysis
 - Functional testing
- AVV clone can now be used to re-infect more HEK293 cells - mass production
- Standard sterile cell line handling & transient transfection*



AVV - purification

- Collect dying infected HEK293 cells (*cpe* = cytopathic effect)
- Rupture plasma membranes to free viral particles (freeze-thaw; ultrasonication)

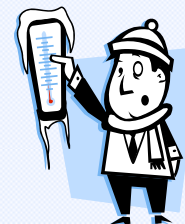


- CsCl ultracentrifugation gradient (2x)
- De-salting column to remove Cs



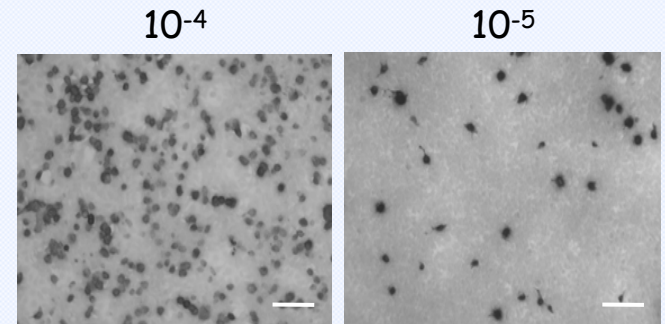
- Sterile filter
- Aliquot, snap-freeze in liquid N₂
- Storage: -80°C

*Alternative:
commercially available
adenoviral vector purification kit*



AVV - titration

- Infect HEK293 cells with serial dilutions of AVV
 - wait 2 days for AVV proliferation to start but not spread (no *cpe*)
- Fix cells and carry out immunocytochemistry against hexons (adenoviral capsid protein)
 - Secondary HRP-coupled antibody, followed by DAB reaction



adapted from: Bewig, B. & Schmidt, W. E. (2000) *Biotechniques* 28, 870-873.

AVV - summary

Advantages

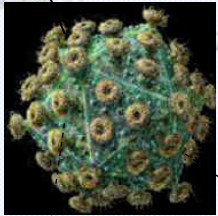
- ability to infect a wide variety of non/dividing cell types: pituitary, thyroid gland, neurones, blood vessel cells
- high efficiency and speed of gene transfer (expression may start already after 12 hours)
- capacity of standard vectors ~ 8kB (there are large capacity "gut-less" adenoviral vectors but they are very difficult to make)
- easy to produce in large quantities
- high titres up to 10^{12} TU/ml (standard types)

Disadvantages

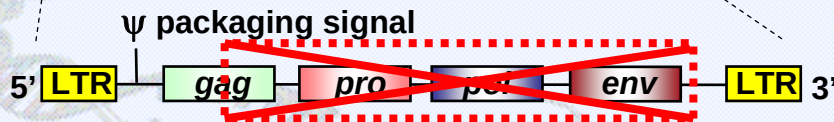
- immune response to viral capsid proteins - especially problematic in peripheral tissues
- Lack of integration + immune response = transient expression (re-administration may be needed)

LVV - background

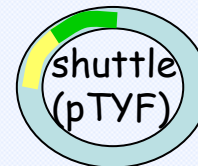
Envelope (coat)



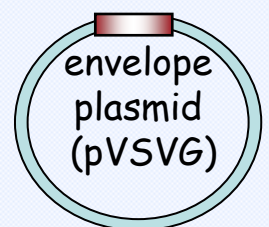
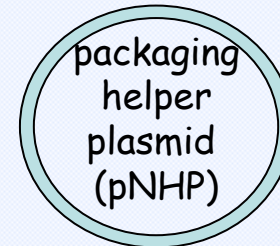
- Derived from HIV, i.e. enveloped, single-stranded RNA viruses
- Pseudotyped with vesicular stomatitis virus (VSVG) coat for receptor-independent transduction
- reverse transcription followed by integration of DNA into host genome



wildtype viral genome

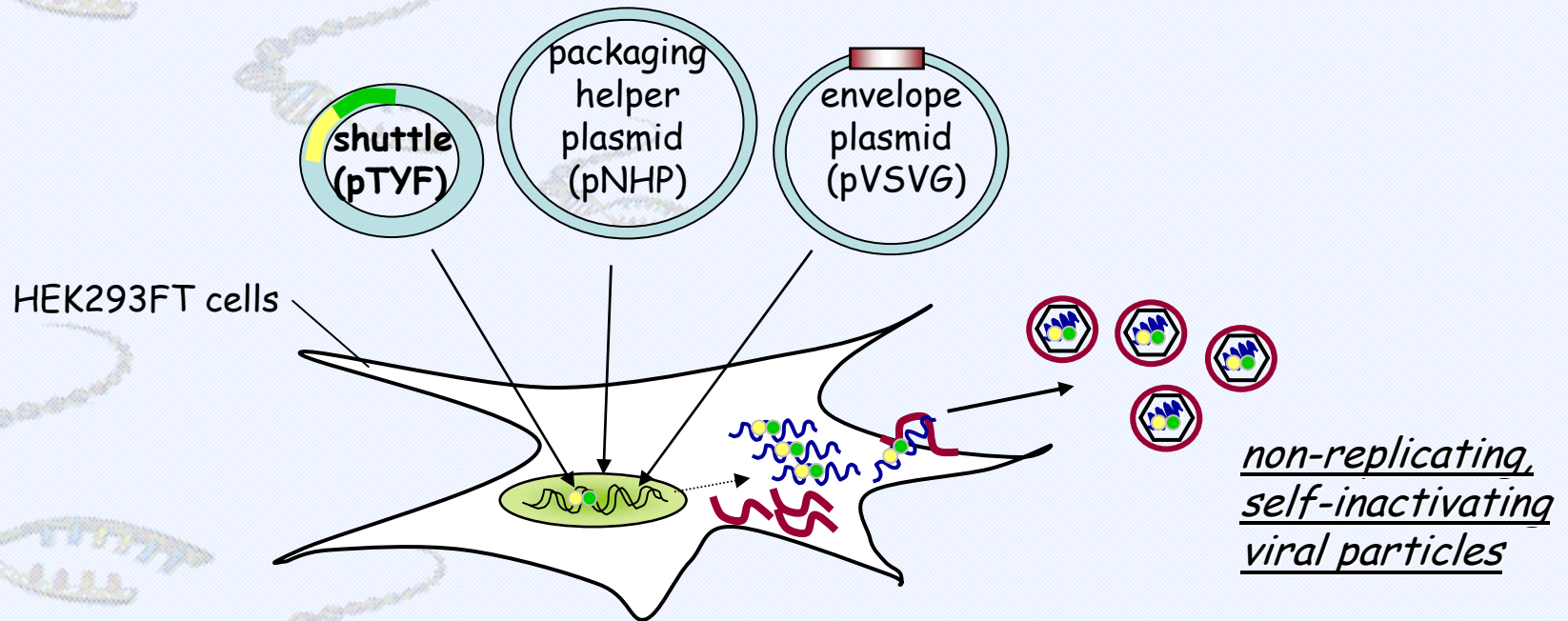


7.5 kb +
transgene
cassette



LVV - construction

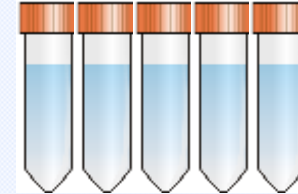
- Co-transfection of shuttle & helper plasmids into HEK293FT cells



- Secretion of viral particles into media (no cell death)
- *Standard sterile cell line handling & transient transfection*

LVV - purification

- Collect media containing LVV at optimal time window



- Concentrate LVV - pelleting by ultra-centrifugation (iodixanol 2x)



*Alternative:
commercially available
lentiviral vector purification kit*

- Aliquot, snap-freeze in liquid N₂
- Storage: -80°C



LVV - titration

- mix into your LVV-containing media a small fraction of PLAP-expressing LVV, and purify together
- infect TE671 cells with serial dilutions of LVV
 - wait 3 days for transgene expression
- fix cells and incubate at 75°C (*this kills all alkaline phosphatase except for PLAP*)
- stain for PLAP activity
- alternatively, immuno-cytochemical titration can be carried out



LVV - summary

Advantages

- can transfect and express genes in a wide range of both dividing and non-dividing cells including neurones *in vivo*
- capacity up to 12 kB (?) - can take complicated constructs
- integrate into host chromosomes - expression more stable
- lack of immunogenic viral proteins - no immune aggression against transduced cells - expression more stable
- easy and quick to produce

Disadvantages

- cannot be obtained in large quantities (for example a typical yield currently is in the range of 50 μ l of 10^9 TU/ml per purification procedure; this compares to 2,5 ml of 10^{11} of adenoviral vectors per procedure).
- risk of mutagenesis if integration occurs into "wrong" host genes (more relevant for gene therapy applications)

The choice between AVV and LVV

<i>criteria:</i>	AVV	LVV
transgene capacity	up to ~8 kb	up to ~12 kb
production time	initially: ~2-4 months later: ~2 weeks *	~2 weeks
titer and quantity	$10^{10} - 10^{11}$ TU/ml 2.5 ml	$10^9 - 10^{10}$ TU/ml 50 μ l
efficient transduction of cell type in exp. system of choice	glia, neurones, other brain cells <i>in vitro</i> and <i>in vivo</i>	prefers neurones <i>in vivo</i> not immunogenic

Teschemacher et al (2005) *Advanced Drug Delivery Reviews* 57:79-93

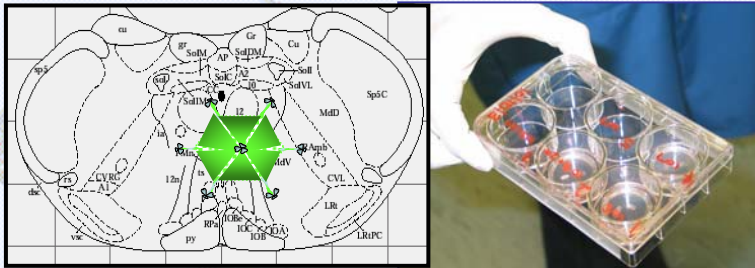
Duale et al (2005) *Experimental Physiology* 90(1):71-78.

Teschemacher et al (2005) *Experimental Physiology* 90(1):61-69.

Transgenesis of brain cells



in vitro: VV transduction in organotypic brainstem slice culture

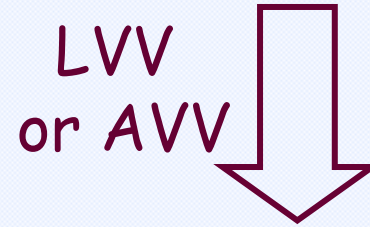


see:

Kasparov et al. (2004) *Prog Biophys Mol Biol* 84:251-277

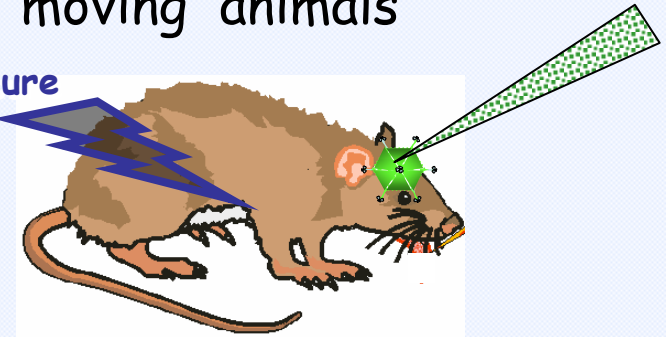
Teschemacher et al (2005) *Advanced Drug Delivery Reviews* 57:79-93.

Teschemacher et al. (2005) *Experimental Physiology* 90(1):61-69.



in vivo: VV microinjections, outcome evaluated in freely moving animals

blood pressure
heart rate



see:

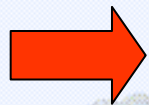
Waki et al. (2003) *J Physiol* 546: 233-242

Duale et al (2005) *Experimental Physiology* 90(1):71-78.

Duale et al (*in press*) *Cardiovascular Research*

Safety issues

- AVV and LVV will not replicate in *your* cells, nor will they survive in the environment
- *however*, in principle, they could deliver transgenes into your cells:



treat as biohazard:



- gloves, lab coat, flow hood, autoclave or incinerate disposable waste
- treat (potentially) contaminated surfaces with viricidal disinfectant (Virkon or bleach)
- observe local genetically modified organisms (GMO) regulations

Standard Equipment !

*cell
culture
handling*

- flow hood for sterile work
- CO² incubator
- centrifuge (50 ml tubes; up to 2000 rmp)
- cell line consumables

*centrifugation
and column
purification*

- rod sonicator (optional)
- ultracentrifuge (or kits)
- -80C freezer
- consumables, e.g. de-salting columns

*Cell line
staining &
counting*

- basic histology facilities
- oven (75C)

Viral Vectors for Cell Type-Specific Transgene Expression in the Brain

- Introduce the transgene into brain cells
 - Make the appropriate delivery vehicle:
 - Adenoviral vector (AVV)
 - Lentiviral vector (LVV)
- Control expression to the desired cell type
 - Choice of Promoter, e.g.:
 - "constitutive" (hCMV)
 - "pan-neuronal" (Synapsin, EF1a, Ta1, NSE)
 - neuronal phenotype-specific:
 - Derived from endogenous promoter (GAD67(3.7))
 - Rational design (PRSx8)
 - activity-dependent (mFOS)
 - glia-specific
 - endothelial-specific



transgene cassette



Targeting transgene expression to a specific cell type



- What is special or unique about gene expression in this cell type? For example:
 - Noradrenergic neurones express dopamine- β -hydroxylase (DBH)
 - GABAergic neurones express glutamate decarboxylase GAD65 or GAD67)
 - Serotonergic neurones express tryptophan hydroxylase (TPH)
 - Many neurones express neuronal markers, e.g. synapsin (Syn), neurone-specific enolase (NSE), etc.
 - Astrocytes express glial fibrillary acidic protein (GFAP)
- Make use of the endogenous transcription machinery of the cell type: to express your transgene, 
 - use a short version of an endogenously active promoter, e.g. GAD67(3.7)- or GFAP-promoters
 - be clever and design a promoter that is switched on by endogenous transcription factors, e.g. PRSx8

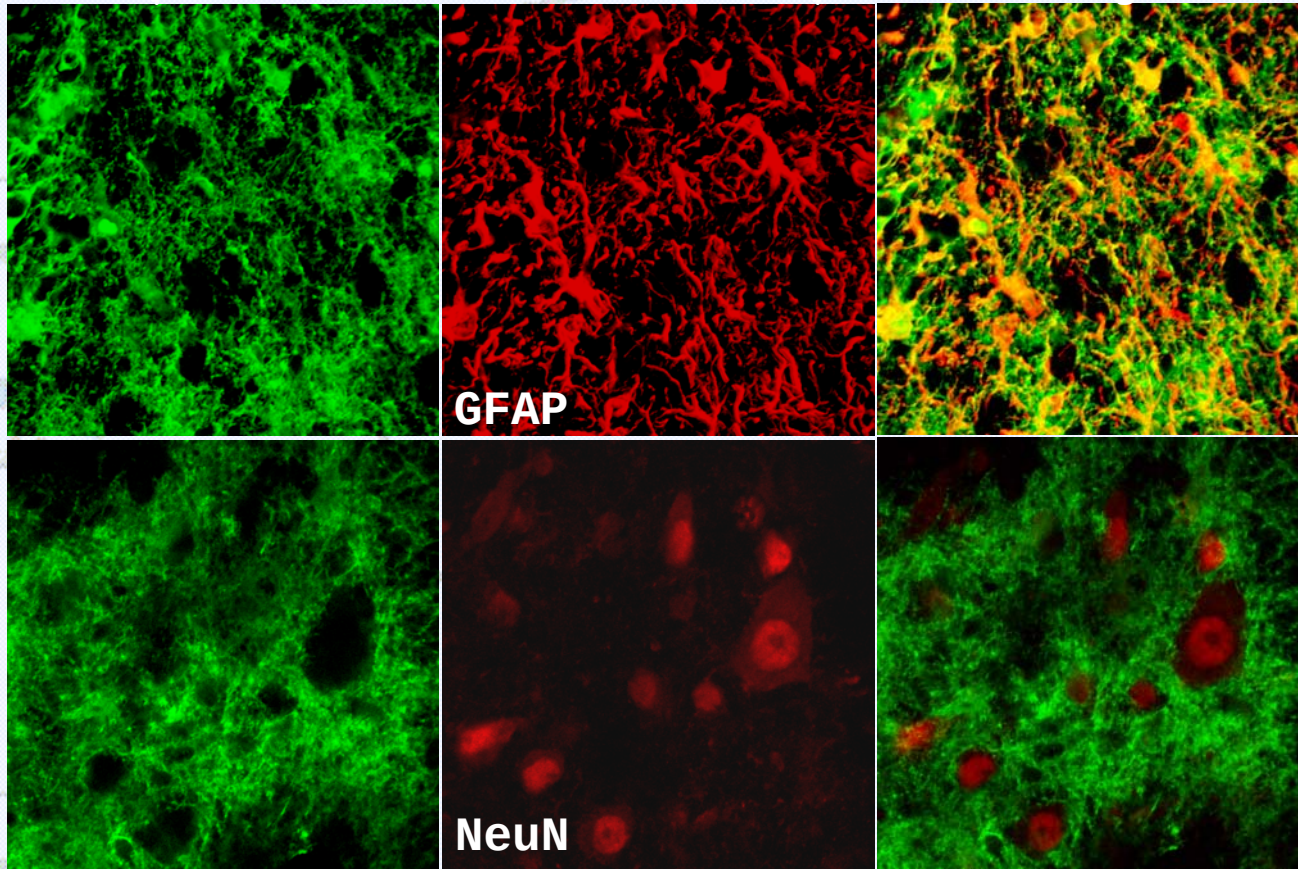
Example 1: Selective transgene expression in astroglia

superGFAP enhanced green fluorescent protein (EGFP)

LVV-superGFAP-EGFP

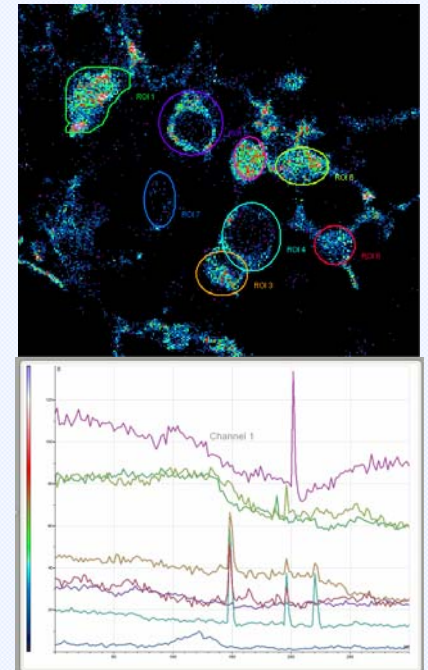
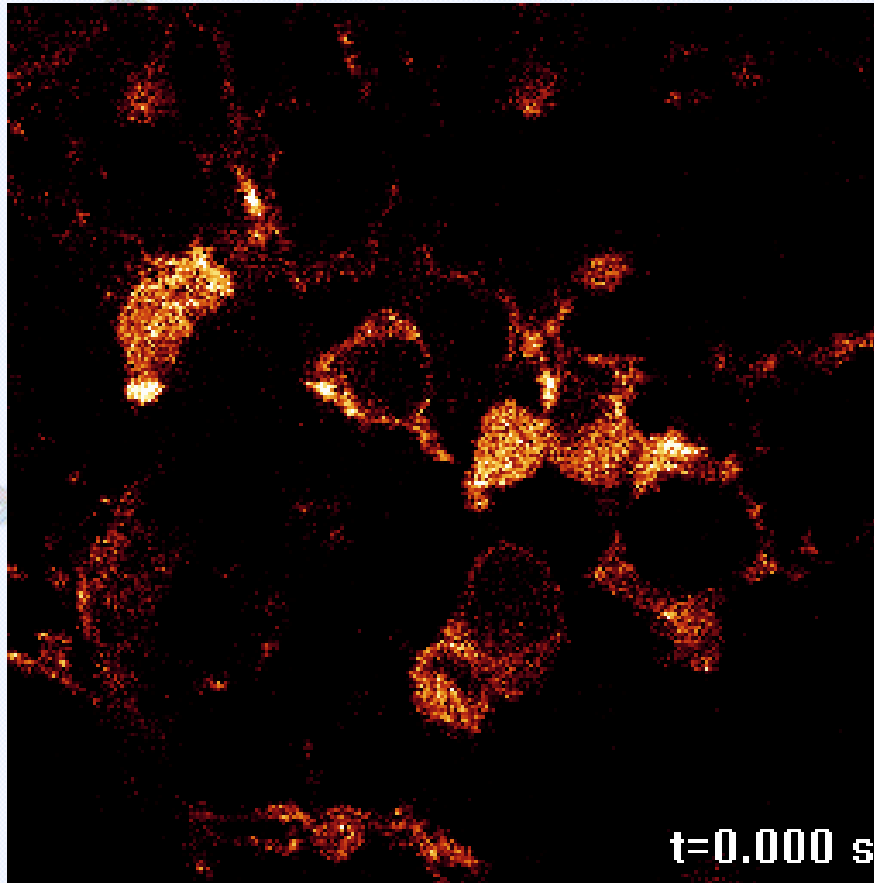
immunoreactivity

merge



Example 1: Selective transgene expression in astroglia

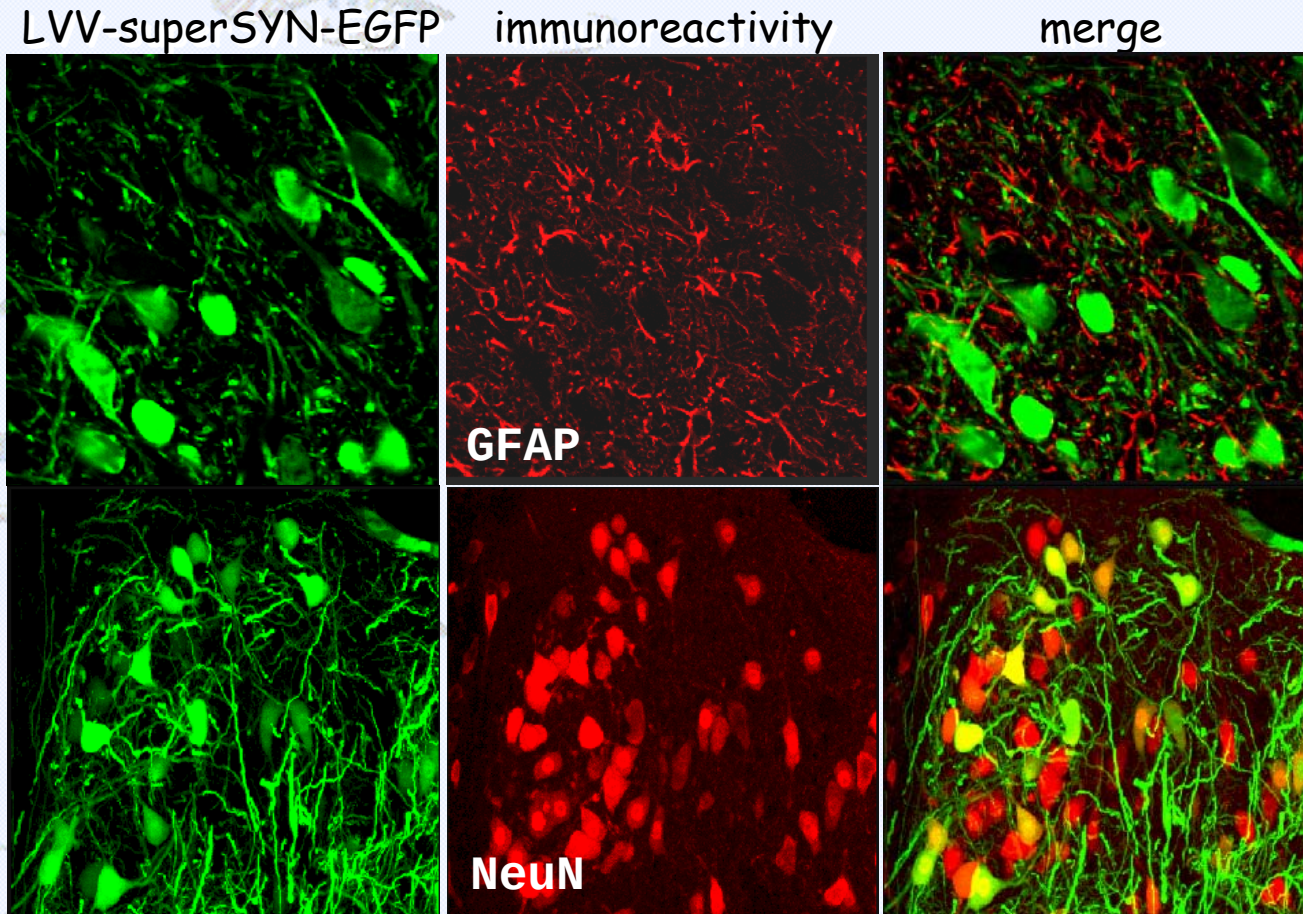
superGFAP $\rightarrow$ Ca^{2+} -sensitive fluorescent protein (cps2r1)



Souslova, E.A. et al, BMC Biotechnology, 2007, 7, 37

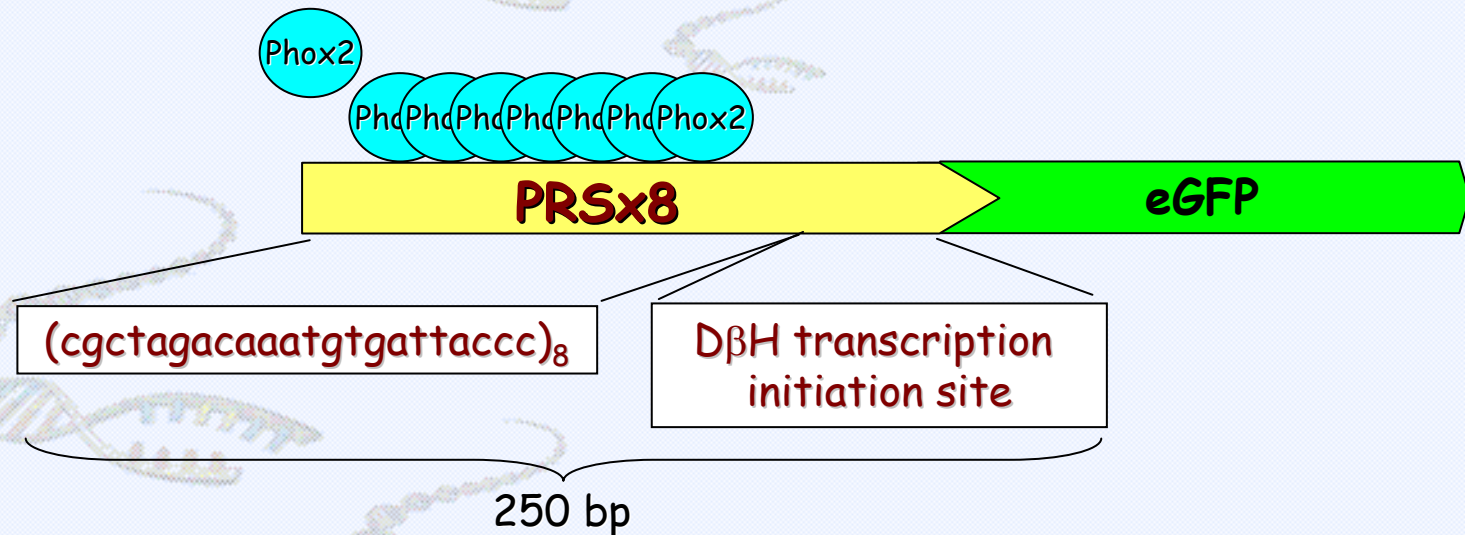
Example 2: Selective transgene expression in neurones

superSYN enhanced green fluorescent protein (EGFP)



Example 3: Selective transgene expression in noradrenergic neurones

➡ In nor/adrenergic neurones, the transcription factor Phox2 is abundantly expressed.

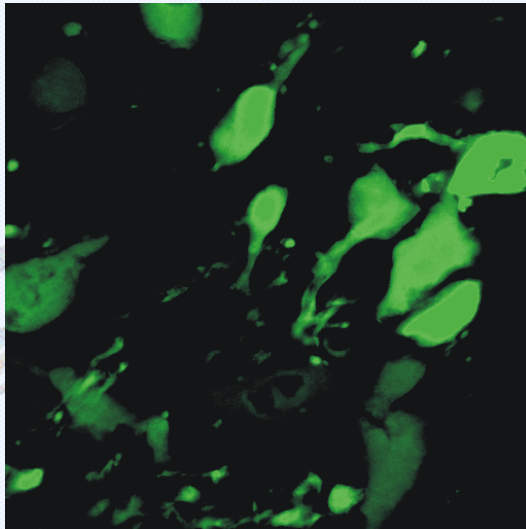


➡ PRSx8 is a short, powerful and specific artificial promoter for VV transgenesis

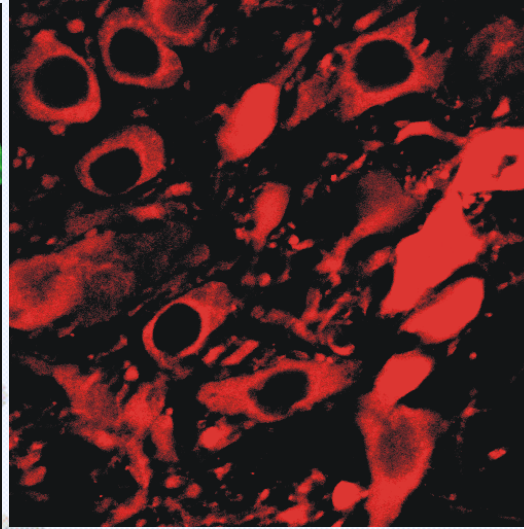
Example 3: Selective transgene expression in noradrenergic neurones

locus coeruleus (A6):

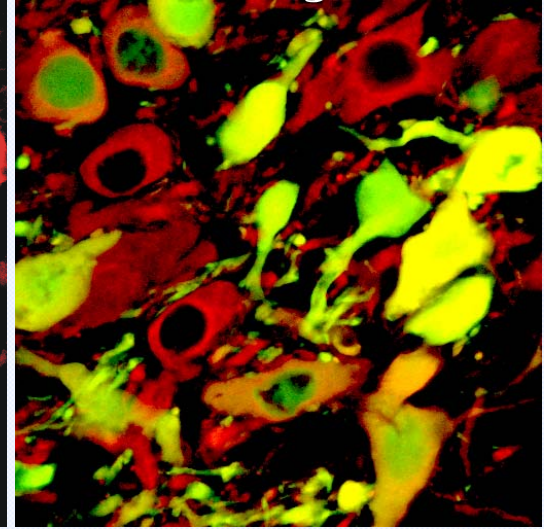
AVV-PRSx8-EGFP



D β H-IR

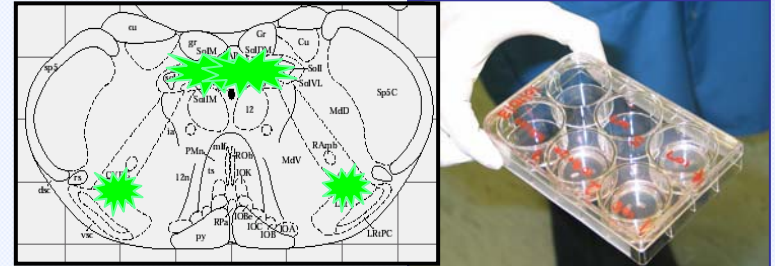
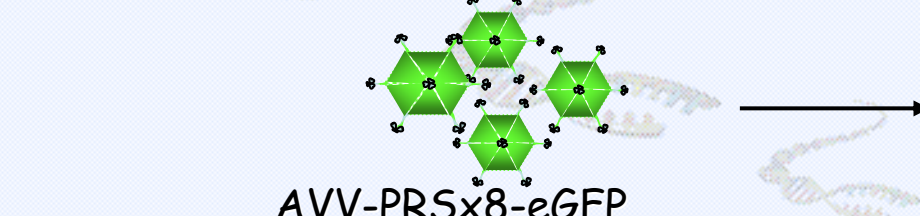


merge

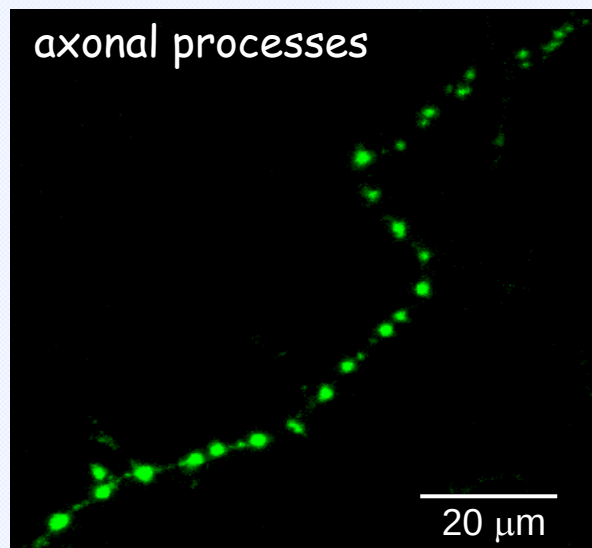
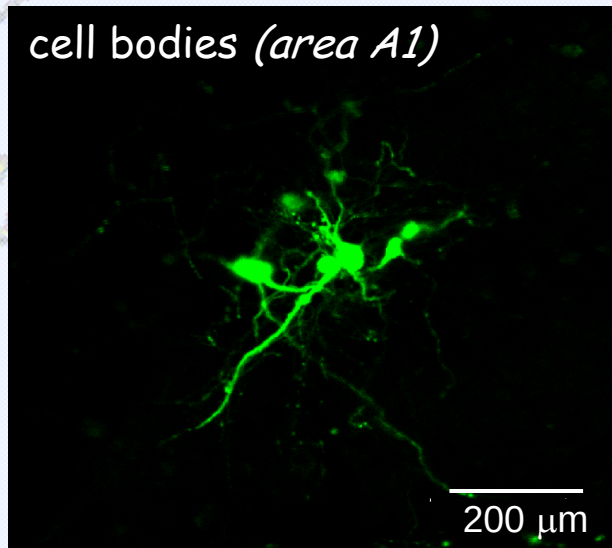


... while other 'constitutive' or 'neurone-specific' promoters (hCMV, SYN, Ta1-EGFP, NSE) are inactive in noradrenergic neurones!

expression in



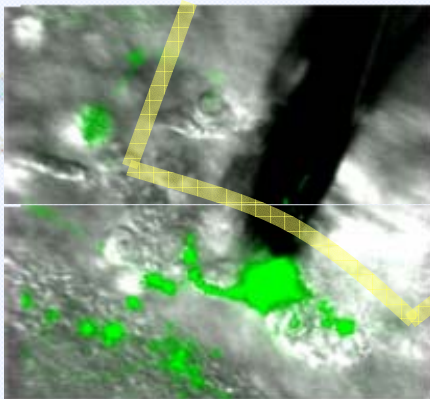
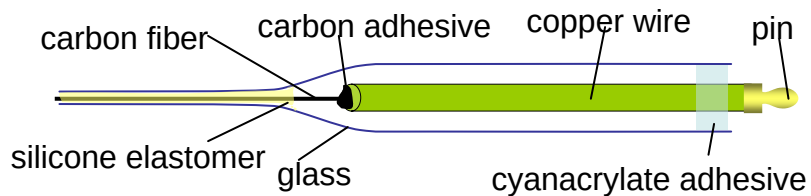
organotypic brainstem slice culture



Teschemacher et al (2005) *Advanced Drug Delivery Reviews* 57:79-93
Teschemacher et al (2005) *Experimental Physiology* 90(1):61-69.

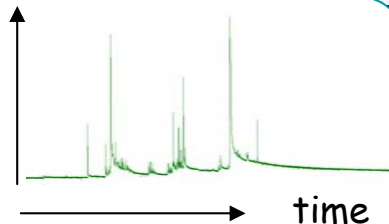
Example 3: Noradrenaline release from brainstem neurones in vitro: Micro-amperometry

Carbon Fibre Electrode:



amplifier
+800 mV

NA
oxidation

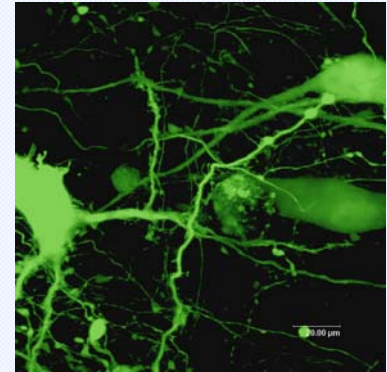
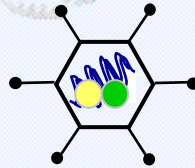
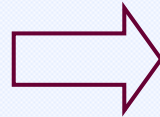


Teschemacher (2005) *Autonomic Neuroscience: Basic and Clinical* 117:1-8.
Chiti & Teschemacher (2007) *FASEB Journal* 21(10):2540-2550

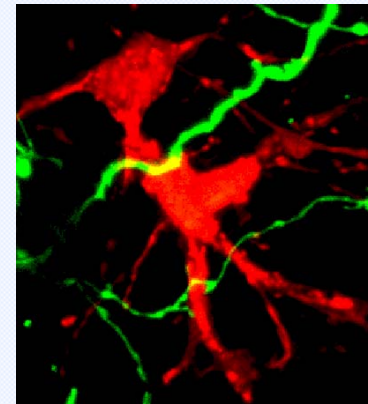
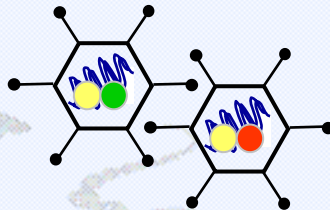
Example 4: Co-expression of cell-specific transgenes for study of cell-cell interactions



Green Fluorescent Protein



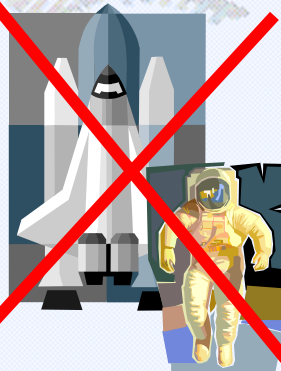
Red Fluorescent Protein



Conclusions

- AVV and LVV are useful tools for cell-specific targeting of brain cells
 - to visualise them for cellular studies (electrophysiology / -chemistry, morphology)
 - To alter their function and investigate their roles in physiology *in vivo*
- They are easy to use and require only a little additional equipment and training

*It's not
rocket
science*



CAN DO IT!

Contacts:

<http://www.bris.ac.uk/Depts/Physiology/Staff/Pysk/virallab/index.htm>
Anja.Teschemacher@bristol.ac.uk; Sergey.Kasparov@bristol.ac.uk

Useful terms:

Gene expression - the process of formation of messenger RNA (mRNA) of a DNA template which then is translated into the sequence of amino acids at the ribosome to make proteins

Transgene - a foreign gene introduced into the cell (for example by a viral vector)

Transduction - process of delivery of a foreign gene into the target cell

In order to make a clear distinction between our constructs and wild type virulent viruses we use the term "vectors"

Expression cassette - a piece of DNA containing elements (promoter, coding part and polyadenylation signal) necessary for expression of a transgene

Acknowledgements

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- Jeff Croker
- Mohan Raizada, Gainesville
- Kwang-Soo Kim, Boston



**British Heart Foundation
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BBSRC
Wellcome Trust
Physiological Society**